



Department of Health and Human Services
Health and Environmental Testing Laboratory
47 Independence Drive
12 State House Station
Augusta, Maine 04333-0012
Tel: (207) 287-2727; Fax: (207) 287-6832
TTY: 1-800-606-0215

Zaire ebolavirus Viral Hemorrhagic Fever Laboratory Submission Information Sheet

Reporting of suspect case to Maine CDC	<ul style="list-style-type: none">•Yes. This is a Notifiable Condition. If you have not already done so, please report suspect or confirmed cases to the Maine CDC via the disease reporting line: 1-800-821-5821 (24hrs/day 7 days/week.)
Required To Submit Laboratory Specimen	<ul style="list-style-type: none">•NO
Required Information	<ul style="list-style-type: none">•Information on requisition must include: suspected organism if known or required, patient name, DOB, date of collection, specimen source or type, submitter name and contact information.•Important: all specimens must be labeled with patient name and be accompanied by a HETL requisition.
Specimen Requirements	<ul style="list-style-type: none">•see page 2
Collection Instructions	<ul style="list-style-type: none">•see page 2
Turn Around Time	<ul style="list-style-type: none">•see page 2
Unacceptable Conditions	<ul style="list-style-type: none">•Specimens received without a form and clinical details or with insufficient material to analyze.•Specimens in poorly labeled or leaking containers.•Do not use wooden shaft or calcium alginate swabs for Influenza PCR.•<u>Samples stored at inappropriate temperatures.</u>
Results	<ul style="list-style-type: none">•Results reported will depend upon the testing performed•Results will only be reported to the submitter listed on the requisition
Laboratory Testing Fee	<ul style="list-style-type: none">•All fees associated with testing will be waived by HETL as long there is prior approval from Federal and Maine CDC.•\$0
Applicable CPT Codes	<ul style="list-style-type: none">•87502, 87503 (Influenza PCR)•87798 (Ebola PCR)•87040 (bacterial culture)
Additional Information	<ul style="list-style-type: none">•Bacteriology at 207-287-1704•Virology, Respiratory, Arbovirus, Serology, and STI at 207-287-1722•Molecular Biology, Sequencing, and Foodborne at 207-287-5769•Mycobacterium and Rabies at 207-287-1706



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Test	Specimen Source	Volume	Specimen Container	Storage Conditions	Sample Stability	Turn Around Time	Shipping
Influenza Real-Time PCR	Preferred: Nasopharyngeal swab/Throat swab in viral transport media, Nasal wash Other Acceptable specimens: Lower respiratory	Minimum 1.0ml synthetic-tipped swabs with plastic shaft only	viral transport media	2 – 8 °C	6 hrs	< 8hr	IATA Category A
Malaria PCR	Whole Blood	Minimum 0.5 ml	EDTA (purple top) tubes	2 – 8 °C	6 hrs	< 8hr	IATA Category A
*Zaire ebolavirus Real-Time PCR	Whole Blood	Minimum 0.5 ml	EDTA (purple top) tubes	2 – 8 °C	24hrs	< 8hr	IATA Category A
Blood Culture	Whole Blood	See mfg protocol	Facility Specific Blood Culture Bottle	Room Temperature	6 hrs	Variable, Maximum of 7 days	IATA Category A

***Collecting Specimens for Ebola Real-Time PCR Testing**

1. Specimens should be obtained when a patient meets the criteria for person under investigation (PUI) including patients with clinical signs, symptoms, and epidemiologic risk factors for Ebola virus disease. If the first specimen is obtained 1-3 days after the onset of symptoms and tests negative and the patient remains symptomatic without another diagnosis, a later specimen is needed to rule-out Ebola virus infection.
2. Staff who collect specimens from PUIs should wear appropriate PPE and should refer to Guidance on Personal Protective Equipment To Be Used by Healthcare Workers During Management of Patients with Ebola Virus in U.S. Hospitals, Including Procedures for Putting On (Donning) and Removing (Doffing).
3. For adults, a minimum volume of 4 mL whole blood is preferable. For pediatric samples, a minimum of 1 mL whole blood should be collected in pediatric-sized collection tubes. Blood must be collected in plastic collection tubes. Do not transport or ship specimens in glass containers or in heparinized tubes.
4. Whole blood preserved with EDTA is preferred, but whole blood preserved with sodium polyanethole sulfonate, citrate or with clot activator is also acceptable.
5. Do not separate and remove serum or plasma from the primary collection container.
6. Specimens should be packaged and transported at 2°–8°C with cold-packs to the final testing destination.